



# Determination of Polystyrene Microplastic in Soil by Pyrolysis – Gas Chromatography – Mass Spectrometry (pyr-GC-MS)

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## ABSTRACT

Pyrolysis-gas chromatography-mass spectrometry (pyr-GC-MS) is emerging as a promising alternative to the detection and quantification of microplastic pollution. For a robust quantification it is essential to improve our understanding of interferences in the pyrolysis of microplastics. Here we investigate the effects of different soil matrices, mainly differing by their organic carbon content ( $C_{org}$ , 1.0–13.6%), and of the polymer Mw on the pyr-GC-MS analysis of polystyrene (PS) microplastics. In addition, we evaluated the effectiveness of adding poly(4-fluorostyrene) (PSF) as internal standard to circumvent the matrix effects. The three main markers of PS pyrolysis, i.e., styrene, styrene-dimer and styrene-trimer, were monitored. The ratio between the dimer and the trimer significantly varied between the matrices and tended to decrease with the increasing of the  $C_{org}$  in the soil, mainly due to increased trimer formation. A strong matrix effect affected the slope of the calibration curves by 2 to 8-fold and was correlated with the  $C_{org}$  in the soils. This effect was mitigated when the areas of the markers were normalized by the area of the corresponding marker of PSF. PS of low Mw (Mw 35,000) presented a reduced formation of the three markers compared to PS of high Mw (Mw 400,000), and styrene-dimer was proportionally less formed than the other two markers. Differences in the slopes of calibration curves depended on the marker chosen, highlighting the relevance of selecting the pyrolysis marker in the quantification of microplastics using pyr-GC-MS.

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Polystyrene microplastic; pyrolysates; pyrolysis – gas chromatography – mass spectrometry (pyr-GC-MS); soil analysis

## Introduction

The pervasiveness of microplastic pollution in the environment, i.e., plastic particles smaller than 5 mm, has been followed by an urgent need for analytical methodologies to detect and quantify this pollutant. Soils have been recognized as an important sink of microplastics (Hurley and Nizzetto 2018), which can alter soil physical properties and

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51 cause decreases in plant height, root biomass, and reproduction rate of soil animals  
52 (Zhang et al. 2022). The current methods to detect microplastics in environmental sam-  
53 ples mainly focus on extensive protocols to reduce the soil organic and inorganic con-  
54 tent to facilitate the microplastic identification (Pinto da Costa et al. 2019; La Nasa  
55 et al. 2020; Thomas et al. 2020). Particles are commonly individually analyzed by vibra-  
56 tional spectroscopy (infrared and Raman) (Käppler et al. 2016; Veerasingam et al. 2021)  
57 and/or the bulk sample is processed for the detection of microplastics, as in mass spec-  
58 trometry-based methods (Dierkes et al. 2019; Okoffo et al. 2020; Velimirovic et al.  
59 2021). Vibrational spectroscopy allows the identification of particles composition, their  
60 physical properties, such as shape and color, and provides the number of particles in a  
61 sample. However, the limit of detection of few micrometers, and the intense sample pre-  
62 treatment in order to better isolate the particles are considered main disadvantages of  
63 these techniques (Renner, Schmidt, and Schram 2018). Thermo-analytical methods, as  
64 pyrolysis coupled to gas chromatography and mass spectrometry (pyr-GC-MS), have  
65 been emerging as a potential alternative for the analysis of microplastics in complex  
66 matrices, such as soils (Peñalver et al. 2020). These methods may shorten the analysis  
67 time due to less need of sample clean-up, potentially allowing a direct injection into the  
68 equipment with reduced extraction steps (Dümichen et al. 2017; Picó and Barceló 2020;  
69 Yakovenko, Carvalho, and ter Halle 2020).

72 A critical step in pyr-GC-MS is the controlled and reproducible pyrolysis of a sample,  
73 which leads to the formation of volatile pyrolysates (Sam 2019). Among these pyroly-  
74 sates, specific markers of the polymers composing microplastics are sought (Ohtani  
75 et al. 1990; Tsuge, Ohtani, and Watanabe 2011). Therefore, the detection of microplastics  
76 is performed indirectly, and the presence of one or more markers is extrapolated to  
77 the presence of a polymer. The pyrolysis of polystyrene (PS), for instance, mainly gener-  
78 ates three markers: the monomer (styrene), the styrene-dimer (3-buten-1,3-diyldiben-  
79 zene), and the styrene-trimer (5-hexen-1,3,5-triyltribenzene) (Tsuge, Ohtani, and  
80 Watanabe 2011; La Nasa et al. 2020). The styrene is commonly the most abundant frag-  
81 ment; however it can also be formed during the pyrolysis of classical soil organic matter  
82 constituents, including lignin, and other synthetic polymers, such as polyethylene ter-  
83 ephthalate (PET) and acrylonitrile butadiene styrene (Saiz-Jimenez and De Leeuw 1986;  
84 Hempfling and Schulten 1990; Dümichen et al. 2015; Dziwiński, Iłowska, and Gniady  
85 2018). Therefore, more specific markers of polystyrene pyrolysis, such as the styrene-  
86 dimer and trimer, are usually screened for the presence of polystyrene in pyr-GC-MS  
87 analyses.

89 The quantification of microplastics in environmental samples by pyr-GC-MS is  
90 mainly based on external calibration curves, resulting in mass estimates. These curves  
91 are established by adding microplastic directly into the pyrolysis system or into a  
92 matrix, which usually differs from the one being analyzed (Fischer and Scholz-Böttcher  
93 2017; Ribeiro et al. 2020; Leslie et al. 2022). However, the pyrolysis of microplastics can  
94 be strongly affected by a matrix effect, and different slopes between curves have been  
95 attributed to an effect of residual mineral components of soil matrices, such as clays  
96 (Fabbri, Trombini, and Vassura 1998; Bouzid et al. 2022). In addition, organic compo-  
97 nents might lead to a variety of volatile pyrolysis products, directly interfering with the  
98 specific markers and affecting the quantification of microplastics (Fischer and Scholz-  
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101 Böttcher 2017; Picó and Barceló 2020). To overcome the effects of complex matrices  
102 in microplastic quantification, the use of an internal standard (ISTD) is recommended  
103 (Unice, Kreider, and Panko 2012), which should ideally mimic the effects suffered by  
104 the target analyte during pyr-GC-MS analysis. Different ISTD have been tested, includ-  
105 ing deuterated polystyrene (PS<sub>D5</sub>) (Dierkes et al. 2019; Fischer and Scholz-Böttcher  
106 2019; Rødland et al. 2020; Scherer et al. 2020). Recently, extensive matrix-dependent H-  
107 D exchanges reactions of PS<sub>D5</sub> have been demonstrated, and the use of poly(4-fluorosty-  
108 tyrene) (PSF) has been preferred (Lauschke et al. 2021). The effectiveness of using PSF  
109 as ISTD to increase the accuracy of microplastic quantification in the analysis of com-  
110 plex matrices using pyr-GC-MS remains to be investigated.

111 The use of calibration curves built with pyrolysis markers of virgin microplastic  
112 standards is under the assumption of a similar pyrolytic behavior between this reference  
113 material and the microplastic in the environment. However, changes in the molecular  
114 structure of the microplastic, such as appearance of carbonyl groups due to an oxidation  
115 process, are shown to affect the formation of its markers during pyrolysis (Ainali,  
116 Bikiaris, and Lambropoulou 2021a; Toapanta et al. 2021). Moreover, polymer molecular  
117 weights (Mw) have already been highlighted as an important determinant of pyrolysis  
118 yields (Audisio and Bertini 1992; Guo et al. 2017; Park and Lee 2021). Styrene yield, for  
119 instance, was shown to increase with the Mw of polystyrene (Bouster, Vermande, and  
120 Veron 1989). In addition, microplastics might present diverse patterns of Mw distribu-  
121 tion (ter Halle et al. 2017), likely differing from the virgin polymers used to build cali-  
122 bration curves. For reliable estimates of microplastic in environmental samples, it is  
123 crucial to understand the effect of Mw on pyr-GC-MS analysis and the consequences  
124 on microplastic quantification.

125 In this study, we aim to investigate the effects of the matrix and the polymer Mw on  
126 the PS quantification using pyr-GC-MS. First, we assess the effect of different soil matri-  
127 ces, mainly differentiated by their organic carbon content, on the pyrolysis of PS. Then,  
128 we verify the effectiveness of PSF as an internal standard to circumvent matrix effects.  
129 Second, we compare the pyrolysate yields from PS of two different Mw and we evaluate  
130 the effect on the quantification of this microplastic. Specifically, we investigate the  
131 changes in the proportion of the main PS markers and the effect on the slope of cali-  
132 bration curves.

## 133 **Methodology**

### 134 **Standards acquisition**

135 Polystyrene microplastics of low Mw (PSLMW, average Mw 35,000, Merck KGaA,  
136 Darmstadt, Germany, 331651, CAS: 9003-53-6) were obtained as pellets and ground in  
137 a microfine grinder (MF10, IKA-Werke GmbH & Co. KG, Germany) at 3000 rpm using  
138 a 1.0 mm sieve. The collected powder was successively sieved through 500, 200, 100,  
139 and 63 µm mesh sieves. Polystyrene microplastics of high Mw (PSHMW, average Mw  
140 400,000) were purchased as a powder with an average particle size of 300 µm (Merck  
141 KGaA, Darmstadt, Germany, 450383, CAS: 9003-53-6). Poly (4-fluorostyrene) (PSF,  
142 average Mw 3,200) was purchased from Polymer Source as the powder (Montreal,  
143 Canada, P43491A).

PSF and PSLMW were dissolved in dichloromethane (Sigma Aldrich, 34856, CAS: 75-09-2) to prepare a solution at  $200 \mu\text{g mL}^{-1}$ . PSHMW was insoluble in the solvents tested (dichloromethane, chloroform, toluene, and ethyl acetate), and no solution could be prepared.

### **Matrices analyzed**

Acid-washed sand,  $\text{SiO}_2$ , (Fisher Chemical, S/0330/65, CAS: 14808-60-7) was heated at  $600^\circ\text{C}$  in a porcelain grail overnight prior to use. Then, the sand (SA) was ground in the Mixer Mill (Retsch MM400) under 20 Hz for 3 min using a single 20 mm diameter zirconium ball. Sand was composed only of sandy texture ( $1000 \text{ mg g}^{-1}$ ), and its carbon organic content was considered as null.

Five soils were obtained from a previous work by Guigue et al. 2014, along with their parameters of organic carbon content (Corg, ranging from  $10 \text{ mg g}^{-1}$  to  $136 \text{ mg g}^{-1}$ , i.e., 1.0–13.6% (w/w)) and other characteristics, such as nitrogen content, carbon/nitrogen ratio, texture and pH. These soils are developed on different source materials, with various vegetation types and cover a wide spectrum of organic matter composition and concentration. Briefly, for each soil, a 5 kg composite of the A-horizon was collected from a soil pit with well-defined horizons. All soils were air-dried, sieved through a 2 mm mesh, and homogenized before further experiments. GL is a Gleyic Luvisol (Corg =  $10 \text{ mg g}^{-1}$ ) collected from an agricultural area in the Burgundy region ( $47^\circ 07' 23''\text{N}$ ;  $5^\circ 05' 08''\text{E}$ ), presenting a massive structure and was bare at the time of sampling. EC is an Eutric Cambisol (Corg =  $25 \text{ mg g}^{-1}$ ) collected in the Burgundy region ( $47^\circ 23' 12''\text{N}$ ;  $4^\circ 39' 19''\text{E}$ ), at the bottom of a valley, at calcareous alluvial deposits surrounded by carbonate rocks. DC is a Dystric Cambisol (Corg =  $30 \text{ mg g}^{-1}$ ) collected in the Burgundy region ( $47^\circ 06' 16''\text{N}$ ;  $4^\circ 25' 55''\text{E}$ ), with vegetation composed mainly of gramineous pasture. EP is an Entic Podzol (C =  $66 \text{ mg g}^{-1}$ ) collected in the Franche-Comté region ( $47^\circ 11' 29''\text{N}$ ;  $5^\circ 33' 50''\text{E}$ ), well-drained and with the presence of a thick, acidic mor-type organic horizon. DA is a dystric Andosol (Corg =  $136 \text{ mg g}^{-1}$ ) collected in the Auvergne region ( $45^\circ 40' 37''\text{N}$ ;  $2^\circ 57' 53''\text{E}$ ), an organic-rich soil characterized by a low bulk density and the presence of allophanes (Guigue et al. 2014).

### **Sample preparation and analysis by pyr-GC-MS**

#### ***PSLMW solution in the presence or absence of matrices***

Calibration curves of PSLMW solution (in dichloromethane,  $200 \mu\text{g mL}^{-1}$ ) were prepared by adding 5 to  $25 \mu\text{L}$  of solution, corresponding to 1 to  $5 \mu\text{g}$  of PSLMW, in pyrolysis cups (Eco-Cup LF, PY1-EC80F, Frontier Lab) already containing  $10 \mu\text{L}$  of PSF (in dichloromethane,  $200 \mu\text{g mL}^{-1}$ ), which was used as an internal standard (ISTD). The pyr-cups were taken under the fume hood at room temperature for 15 min to allow the solvent to evaporate. A total of seven curves were built, with and without the addition of 5 mg of each matrix analyzed, already milled in the Mixer Mill (SA, GL, EC, DC, EP, DA, displayed in the increasing order of Corg). Procedure blanks consisted of the matrix without the addition of PSLMW solution and submitted to the same sample procedure.

### ***Matrices spiked with PSLMW and PSHMW powder***

To build calibration curves, a first sample (solution A) was prepared by the addition of 40 mg of each type of PS, individually, into 4 g of matrix to a final concentration of 10 g kg<sup>-1</sup>. The SA matrix was used for PSLMW (200 – 500 μm fraction), while GL, EC, DC, EP and DA were used for PSHMW. This content was mixed and ground in the mixer mill. Then, a fraction was weighed (Sartorius, SECURE 225D-15, readability = 0.01 mg) and spiked into a matrix to prepare solutions of 1.25, 1.00, and 0.50 g kg<sup>-1</sup>. From these secondary samples, a fraction was weighed to prepare samples of 0.25, 0.10, and 0.05 g kg<sup>-1</sup>, respectively. Finally, each sample was ground by the mixer mill. The final concentrations were calculated considering the weight of sample added and the weight of blank matrix, resulting in small variations in the concentrations at the same calibration level. Procedure blanks consisted of the matrix without the addition of PS and submitted to the same sample procedure.

Approximately 5 mg of each sample with a concentration of 0.05 to 1.25 g kg<sup>-1</sup> and procedure blanks were weighed in a microbalance (Sartorius, CUBIS MSU 6.6S, readability = 0.001 mg) directly into a pyr-cup. The value was recorded for the nearest 0.001 mg. The amount of injected polymer was calculated by the mass of the sample and its concentration. When indicated, this final step was repeated, and it is displayed as replicate.

### ***Contamination control***

To avoid any contamination during the sample preparation procedure, nitrile gloves and cotton lab coats were always worn. The working surface was cleaned with ethanol and deionized water before each procedure, and glassware and metalware were used whenever possible, also rinsed with deionized water before their use. All steps during the procedure were preferably performed under a fume-hood, and sample containers were always covered with aluminum foil to avoid airborne contamination. All pyrolysis cups for pyr-GC-MS were heated at 600 °C in a porcelain grail overnight prior to use. Procedural blanks, composed only by the matrix under study, were submitted to the same procedure as the samples and were further analyzed to check for the presence of sources of contamination.

### ***Pyr-GC-MS analysis***

Samples in pyr-cups were covered with quartz wool (ThermoFisher, 33822200, CAS: 60676-86-0,) and then analyzed by pyrolysis (EGA/Py 3030D, autosampler, FrontierLab) – gas chromatography (GC 7890, Agilent Technologies, Santa Clara, CA USA) – mass spectrometry (MSD 5978B, Agilent Technologies, Santa Clara, CA USA). Pyrolysis was performed at 600 °C for 30 s under helium, followed by a chromatographic separation in an Ultra-Alloy+ – 5 column (30 m × 0.25 mm × 0.25 μm, Frontier Lab), with helium as a carrier gas and oven program at 40 °C for 2 min, 20 °C min<sup>-1</sup> until 320 °C and hold for 6 min. An interface temperature of 300 °C, split of 1:20 and a constant flow of 1 mL min<sup>-1</sup> were settled. The molecules were ionized at the source (EI, 70 eV, 230 °C) and the quadrupole analysis included the full scan (from m/z 45 to 600) and SIM modes. The most intense m/z of the main compounds originated from the



**Table 1.** Markers obtained from the pyr-GC-MS analysis of polystyrene low molecular weight (PSLMW), polystyrene high molecular weight (PSHMW) and poly (4-fluorostyrene) (PSF), and their respective m/z monitored in single ion monitor (SIM) mode and Kovats' retention indices (RI).

Compound	Polymer	m/z	RI
Styrene (monomer)	PSLMW or PSHMW	104	902
3-buten-1,3-diyldibenzene (dimer)		91	1741
5-hexen-1,3,5-triyltribenzene (trimer)		91	2496
4-Fluorostyrene (styrene-F)	PSF	122	908
2,4-di(4-fluorophenyl)but-1-ene (dimer-F <sub>2</sub> )		109	1749
2,4,6-tri(4-fluorophenyl)hex-1-ene (trimer-F <sub>3</sub> )		109	2470

pyrolysis of PSLMW/PSHMW or PSF was monitored (Table 1). The retention time for each compound was defined by the injection of PSLMW and PSF solution and the spectra comparison with the literature (Tsuge, Ohtani, and Watanabe 2011) and with the NIST 14 library (v. 2.3). Kovats' Retention indices (RI) were calculated using the retention times of the n-alkanes determined in the pyrogram of polyethylene under the same pyr-GC-MS conditions (IUPAC 1997).

### Data analysis and visualization

The data from peak integration was obtained with the MassHunter Workstation software (v. B.07.00, Agilent Technologies, Santa Clara, CA USA) and the data analyses were performed at R v.4.2.2 (R Core Team 2022).

The procedural blanks and the instrumental blanks (empty cups) were analyzed to check the presence of the target compounds. Procedural blanks were analyzed in the same sequence of the respective samples, and instrumental blanks were injected at every injection of 5 samples. No peaks of the target compounds were detected in the blanks, except for the styrene in soil samples, indicating that no contamination occurred during sample handling.

The surface area obtained for each marker of PS pyrolysis was always corrected by the weight of sample to obtain the corrected mass of polymer injected in each analysis. Therefore, small variations in calibration levels are expected, without affecting the purposes of this study. Linear regression models were applied to test the relationship between the mass of the polymer and the surface area of the markers. Analysis of variance (ANOVA) tests (stats package v 4.1.2, R Core Team 2022) were computed to estimate the effects of matrix or polymer Mw and the injected polymer mass, or their interaction, on the area of markers, always carried out within a marker type, once different markers are expected to have different responses. In the presence of a significant interaction, post-hoc pairwise comparison tests were performed with emmeans function (emmeans package, v 1.7.2, Lenth et al. 2023).

To analyze changes in the formation rate of pyrolysates against different matrices or polymer Mw, two indicators were explored. The first consisted of the percentage of styrene monomer (Equation 1); dimer or trimer and was applied for samples without matrix or in the presence of sand. In the presence of soils, and therefore of styrene naturally originated from the pyrolysis of these matrices, the formation of dimer and trimer was explored by their ratio (dimer/trimer, or  $R_{D/T}$ ). In the absence of styrene-dimer signal (low PS concentrations), the ratio was not calculated. Peak area ratios can

301 be compared between different samples to allow the identification of different pyrolysis  
302 pattern.

$$303 \text{ Styrene (\%)} = \frac{\text{Area styrene}}{(\text{Area styrene} + \text{Area dimer} + \text{Area trimer})} * 100$$

306 Equation 1. Calculation of the relative percentage of styrene against the three main  
307 markers

308 Differences on these indicators were tested by Wilcoxon or Kruskal-Wallis tests (stats  
309 package, R Core Team 2022). If the Kruskal-Wallis test was significant ( $p < 0.05$ ), a  
310 post-hoc analysis was performed to determine which levels differed from each other by  
311 Dunn test (FSA package, v 0.9.3, Ogle, Doll, and Wheeler 2023). Significant differences  
312 between these indicators, i.e.,  $p < 0.05$ , among different matrices or polymer types are  
313 indicated in each plot as different letters.

314 Correlations between the Corg and the (i)  $R_{D/T}$  and (ii) the slope of calibration curves  
315 were tested using Spearman correlation tests (stats package). Before testing the correl-  
316 ation between the other soil parameters, i.e., nitrogen content and soil texture (sand,  
317 loam, and clay composition, Table S1), and (i) and (ii), we tested the correlation among  
318 soil parameters. The Corg was strongly and positively correlated with nitrogen content  
319 ( $\rho = 0.99$ ,  $p < 0.01$ ), and the sand content was negatively correlated with the loam con-  
320 tent ( $\rho = -0.94$ ,  $p < 0.01$ ). Therefore, sand and clay compositions were the soil parame-  
321 ters further used to test the correlation with (i) and (ii).

322 For the study of the effectiveness of ISTD, the surface area obtained for each marker  
323 of PSF pyrolysis, i.e., styrene-F, dimer-F<sub>2</sub> and trimer-F<sub>3</sub>, was used to normalize the sur-  
324 face area of the equivalent marker of PS, i.e., styrene, dimer and trimer. To analyses  
325 changes in the formation of PFS pyrolysates, the percentage of styrene-F and the ratio  
326 dimer-F<sub>2</sub>/trimer-F<sub>3</sub> ( $R_{DF2/TF3}$ ) were calculated.

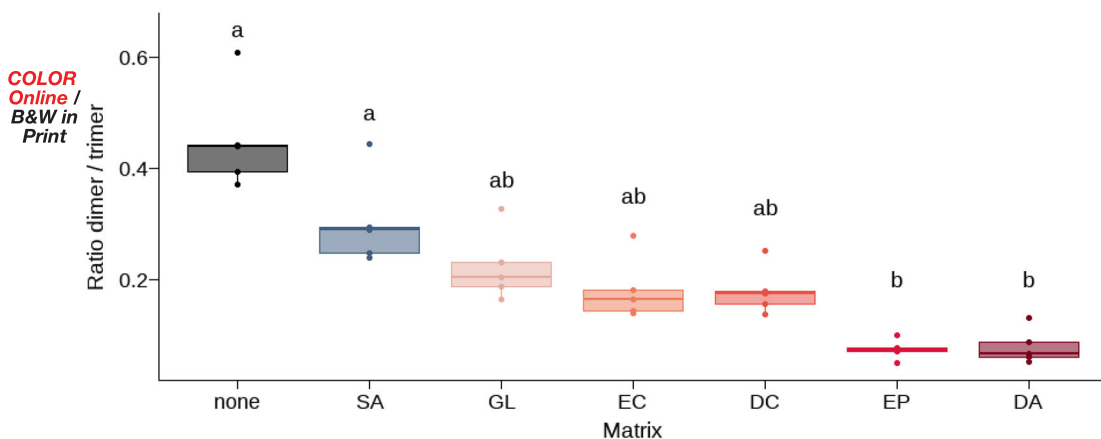
327 The data were plotted using ggplot2 package v 3.3.5 (Wickham et al. 2023). In boxplots,  
328 the lower and upper hinges correspond to the first and third quartiles. The upper whisker  
329 extends from the hinge to the largest value no further than  $1.5 * \text{IQR}$  from the hinge  
330 (where IQR is the inter-quartile range, or distance between the first and third quartiles).  
331 In linear plots, the symbols represent the actual values, and the lines represent the fitted  
332 trend, while shadows around the curve represent the 95% confidence interval.

## 333 Results

### 334 *Effect of matrix upon the pyrolysis of PSLMW*

335 In the procedural blanks, no PS markers were detected for the SA matrix. For blank soil  
336 matrices, a peak of styrene was noticed (Figure S1a) and their surface areas were strongly  
337 correlated with the Corg of the matrices (Spearman,  $\rho = 0.99$ ,  $p < 0.01$ ) (Figure S1b).

338 Pyr-cups spiked with PSLMW solution in the presence or absence of matrices were  
339 analyzed to evaluate their effect on the pyrolysis of this polymer. In the absence of  
340 matrix, the styrene was the major pyrolysate of PSLMW, accounting for 94.5% ( $\pm 0.5$ )  
341 of the three markers, followed by the trimer ( $3.8 \pm 0.5$ ) and the dimer ( $1.7 \pm 0.1$ ) ( $n = 5$ ).  
342 The same profile was observed with the addition of matrix; however, the percentage of  
343 each marker significantly varied between the matrices (Kruskal-Wallis,  $p < 0.01$ ). The



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**Figure 1.** Ratio of dimer/trimer ( $R_{D/T}$ ) from pyr-GC-MS analysis of polystyrene low molecular weight (PSLMW) without or with the presence of matrices (sand (SA), gleyic luvisol (GL), eutric cambisol (EC), dystric cambisol (DC), entic podzol (EP), dystric andosol (DA),  $n = 5$ ). Darker red represents matrices with higher organic carbon content. Different letters indicate significant differences ( $p < 0.05$ ).

370 styrene accounted for 88.8% ( $\pm 1.0$ ), on average, of the three markers for the DA  
371 matrix, while the trimer represented 10.3% ( $\pm 1.2$ ) (Table S1). No correlation was found  
372 between the areas of the three markers. The trimer areas were significantly and posi-  
373 tively correlated with the Corg of matrices (Spearman test,  $\rho = 0.99$ ,  $p < 0.01$ ). No other  
374 significant correlation was found between the marker's areas and the soils parameters.

375 The  $R_{D/T}$  formed during the pyrolysis of PSLMW varied significantly in the presence  
376 of different matrices (Figure 1) and were negatively correlated with the Corg of matrices  
377 (Spearman test,  $\rho = -0.86$ ,  $p < 0.01$ ), notably due to the increased formation of trimer.  
378 No significant correlation was found between the  $R_{D/T}$  and the contents of sand and  
379 clay in the soils (Spearman test,  $p > 0.05$ ). The differences among matrix types were sig-  
380 nificant, even though a variation in the  $R_{D/T}$  with different amounts of polystyrene  
381 pyrolyzed was noticed within each matrix. The outlier ratio value in the upper whisker  
382 within each matrix (Figure 1) corresponds to the smallest injected mass of PSLMW, i.e.,  
383 1  $\mu\text{g}$ , demonstrating the existence of an optimal range for analytical detection.

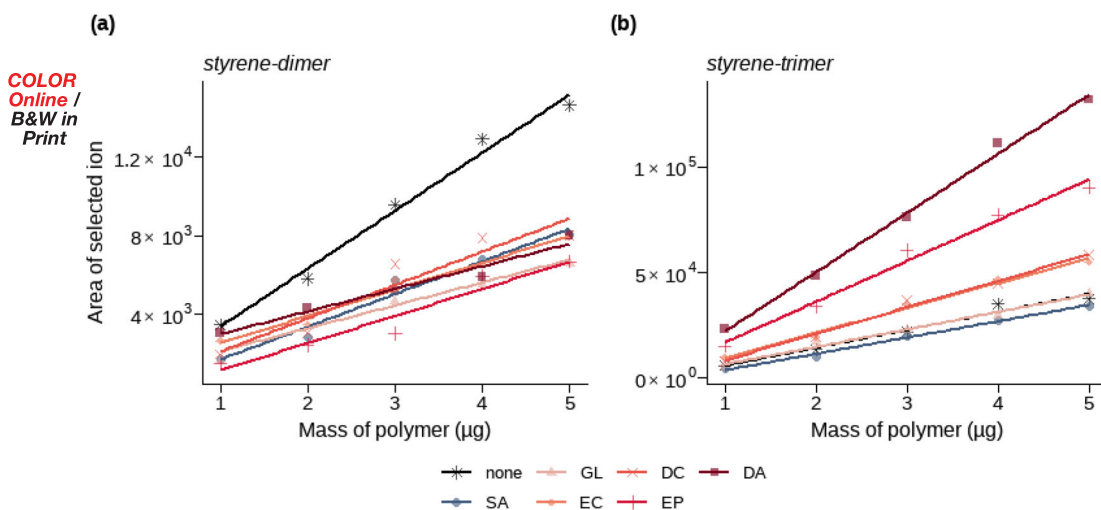
384 Calibration curves built with each marker presented an overall good linearity ( $R^2$   
385 greater than 0.88, Figure 2 and Table S2). Thus, the presence of the matrix had a sig-  
386 nificant and distinct effect on the response of styrene-dimer and trimer to the pyrolysis  
387 of PSLMW (ANOVA,  $p < 0.01$ ) (Figure 2). In the presence of a matrix, the curves for  
388 the styrene-dimer were up to 3-fold less steep (Figure 2a), indicating reduced formation  
389 of this marker. For the styrene-trimer, the differences in slopes were up to 4-fold and  
390 matrix-dependent (Figure 2b), positively and strongly correlated with the Corg of matri-  
391 ces (Spearman test,  $\rho = 1.0$ ,  $p < 0.01$ ). The only significant correlation between the  
392 slopes and the other soil characteristics was found for the slopes of dimer curves and  
393 the sand content (Spearman test,  $\rho = 0.6$ ,  $p < 0.01$ ).

### 396 Attenuation of matrix effect using ISTD

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Pyr-cups spiked with PSF solution in the presence of absence of matrices were analyzed to evaluate the pyrolysis of this polymer. The formation of the three main markers of





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**Figure 2.** Calibration curves of polystyrene markers (a) styrene-dimer and (b) styrene-trimer from polystyrene low molecular weight (PSLMW) solution in the absence or presence of matrix (sand (SA), glycy luvisol (GL), eutric cambisol (EC), dystric cambisol (DC), entic podzol (EP), and dystric andosol (DA)). Darker red represents matrices with higher organic carbon content.

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**Table 2.** Peak area of the three main poly (4-fluorostyrene) (PSF) markers formed during the pyr-GC-MS analysis in the absence or presence of matrix (sand (SA), glycy luvisol (GL), eutric cambisol (EC), dystric cambisol (DC), entic podzol (EP), and dystric andosol (DA),  $n = 5$ ).

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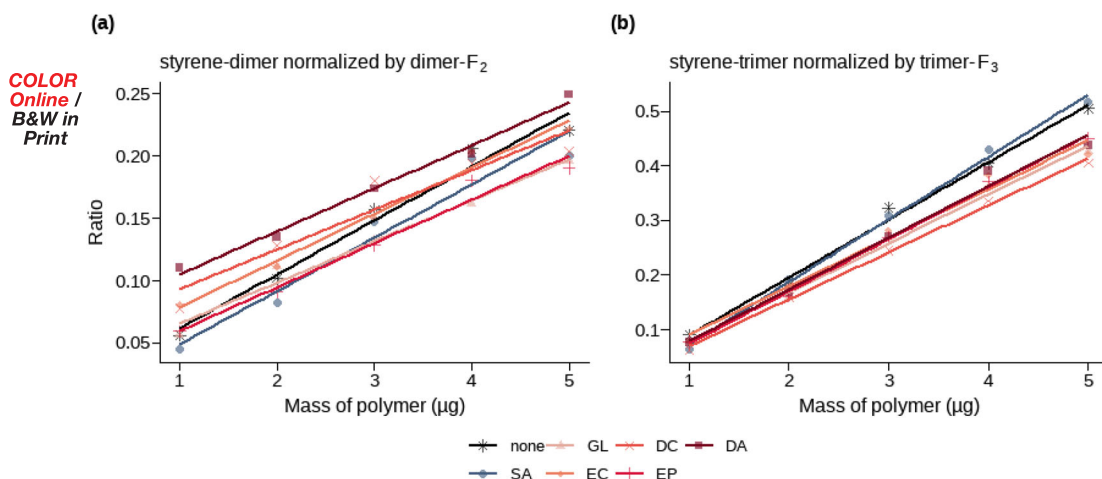
Matrix	Average Styrene-F $\pm$ SD ( $\times 10^6$ )	Average Dimer-F <sub>2</sub> $\pm$ SD ( $\times 10^4$ )	Average Trimer-F <sub>3</sub> $\pm$ SD ( $\times 10^4$ )	Average ratio dimer-F <sub>2</sub> /trimer-F <sub>3</sub> $\pm$ SD
None	1.23 $\pm$ 0.10	6.17 $\pm$ 0.34	7.24 $\pm$ 0.87	0.86 $\pm$ 0.10
SA	1.13 $\pm$ 0.12	3.72 $\pm$ 0.27	6.04 $\pm$ 0.61	0.62 $\pm$ 0.05
GL	1.29 $\pm$ 0.03	3.32 $\pm$ 0.28	8.54 $\pm$ 1.59	0.40 $\pm$ 0.05
EC	1.30 $\pm$ 0.06	3.40 $\pm$ 0.30	12.0 $\pm$ 0.86	0.28 $\pm$ 0.02
DC	1.33 $\pm$ 0.13	3.30 $\pm$ 0.80	13.4 $\pm$ 1.24	0.24 $\pm$ 0.05
EP	1.70 $\pm$ 0.08	2.86 $\pm$ 0.51	20.7 $\pm$ 1.15	0.14 $\pm$ 0.03
DA	1.66 $\pm$ 0.05	3.01 $\pm$ 0.19	29.2 $\pm$ 0.99	0.10 $\pm$ 0.01

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PSF pyrolysis was strongly impacted by the presence of the matrix (Kruskal-Wallis,  $p < 0.01$ ), as it was observed for the pyrolysis of PSLMW. Dimer-F<sub>2</sub> was gradually less formed from the SA to DA matrix, while the opposite profile was observed for the styrene-F and trimer-F<sub>3</sub> (Table 2). The formation of styrene-F and trimer-F<sub>3</sub> was strongly and positively correlated with the Corg of matrices (Spearman,  $\rho = 0.80$  and  $\rho = 0.96$ , respectively,  $p < 0.01$ ). No correlation was found between the areas of styrene-F and dimer-F<sub>2</sub>, while the former was positively correlated with trimer-F<sub>3</sub> areas (Spearman,  $\rho = 0.79$ ,  $p > 0.01$ ). No other significant correlation was found between the marker's formation and other soils parameters.

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The  $R_{DF_2/TF_3}$  from PSF pyrolysis (Figure S2) showed a behavior similar to the  $R_{D/T}$  from PSLMW in the presence of different matrices (Figure 1), and were also negatively and strongly correlated with Corg of matrices (Spearman,  $\rho = -0.97$ ,  $p < 0.01$ ). No other significant and strong correlation was found between the  $R_{DF_2/TF_3}$  and other soils parameters. As for the PSLMW case, the decreasing  $R_{DF_2/TF_3}$  from SA to DA matrix was mainly attributed to the increased formation of the trimer.



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**Figure 3.** Calibration curves of polystyrene markers (a) styrene-dimer (normalized by dimer-F<sub>2</sub>) and (b) styrene-trimer (normalized by trimer-F<sub>3</sub>) from pyr-GC-MS analysis of polystyrene low molecular weight (PSLMW) solution in the presence of different matrices (sand (SA), glyic luvisol (GL), eutric cambisol (EC), dystric cambisol (DC), entic podzol (EP), dystric andosol (DA)). Darker red represents matrices with higher organic carbon content.

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Using the monomer styrene-F to normalize the surface areas of styrene-dimer and trimer did not circumvent the matrix effect (Figure S3a and b, respectively). On the other hand, normalization of styrene-dimer by dimer-F<sub>2</sub> eliminated the matrix effect for styrene-dimer, i.e., the differences in the slopes of the curves (Figure 3a). For the styrene-trimer curves, normalization of styrene-trimer by trimer-F<sub>3</sub> significantly reduced the difference between the slopes to 1.3 fold, although a significant interaction between them and the matrix type was still present (ANOVA,  $p < 0.01$ , Figure 3b). In most cases, the normalization by the ISTD improved the fit of linear models ( $R^2$ ) of the calibration curves (Table S2).

### 483 484 Effect of polymer Mw on the pyrolysis of PS

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For the comparison between the pyrolysis of PSLMW and PSHMW, sand spiked with the respective microplastic powder was used. In general, the signal of PS markers in the pyrolysis of PSLMW was always less intense than those of PSHMW for the same amount of microplastic spiked in sand (Figure S4). Considering the three markers, styrene is proportionally more formed for PSLMW than for PSHMW (Table 3, Wilcoxon test,  $p < 0.01$ ). Conversely, more styrene-dimer was formed for PSHMW than for PSLMW (Table 3, Wilcoxon test,  $p < 0.01$ ), whereas no significant difference was observed in the formation of styrene-trimer (Table 3). The  $R_{D/T}$  was significantly greater for PSHMW (Wilcoxon test,  $p < 0.01$ , Table 3), notably due to the favored formation of dimer.

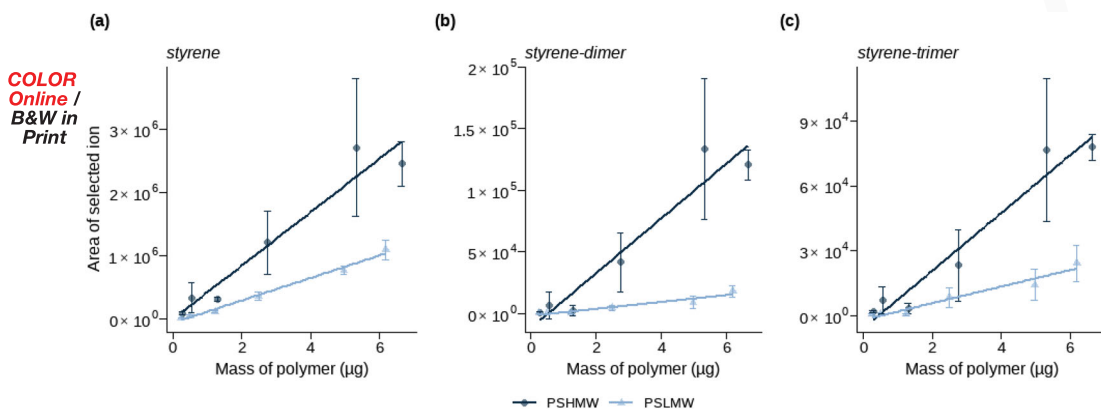
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Calibration curves were built with the three markers obtained from PSHMW or PSLMW spiked in sand ( $R^2$  bigger than 0.78, Figure 4 and Tables S4 and S5). All three markers were detected in the smallest concentration for PSHMW, i.e., 0.05 kg kg<sup>-1</sup>, but the styrene-dimer was only detected in one of the replicates. The styrene-dimer was

**Table 3.** Proportion of the three main polystyrene markers formed during the pyr-GC-MS analyses of polystyrene low molecular weight (PSLMW) or polystyrene high molecular weight (PSHMW) spiked in sand (averaged  $\pm$  standard deviation, SD).

Polymer	Average of styrene (%) $\pm$ SD <sup>(*)</sup>	Average of dimer (%) $\pm$ SD <sup>(*)</sup>	Average of trimer (%) $\pm$ SD	Ratio dimer / trimer $\pm$ SD <sup>(*)</sup>
PSHMW ( <i>n</i> = 12)	94.14 $\pm$ 1.73	3.59 $\pm$ 1.16	2.27 $\pm$ 0.67	1.60 $\pm$ 0.37
PSLMW ( <i>n</i> = 9)	96.72 $\pm$ 0.98	1.29 $\pm$ 0.40	2.00 $\pm$ 0.63	0.65 $\pm$ 0.13

Significant differences ( $p < 0.05$ ) are displayed with an asterisk.



**Figure 4.** Calibration curves of polystyrene markers (a) styrene, (b) styrene-dimer and (c) styrene-trimer from pyr-GC-MS analyses of polystyrene high molecular weight (PSHMW) (dark blue) and polystyrene low molecular weight (PSLMW) (light blue) spiked in sand ( $n = 3$ ).

only consistently detected when the concentration was greater than  $0.25 \text{ g kg}^{-1}$ , equivalent to  $1.25 \text{ µg}$  of polymer (Table S3). The average RSD for styrene areas were higher for PSHMW than PSLMW (35.3% and 19.8%, respectively); however the opposite was noticed for the dimer and trimer (36.9% and 52.6% for PSHMW and 42.8% and 84.9% for PSLMW) (Table S3). The highest relative standard deviation of slopes was found for the curves of trimer from PSLMW (RSD 32.1%,  $n = 3$ , Table S3). The slopes of the curves differed significantly among PS types for all markers (ANOVA,  $p < 0.01$ , Figure 4) from 2 to 8-fold, with the greatest variation for the styrene-dimer (Figure 4b). Using the linear equation obtained from PSHMW to estimate the PS concentration in a sample containing PSLMW lead to underestimates in a 2.5 to 5.3-fold range, depending on the marker chosen (Table S5).

### Analysis of soils spiked with PSHMW

For PSHMW microplastic powder spiked in different matrices and submitted to a milling process, the  $R_{D/T}$  significantly differed between matrix types (Kruskal-Wallis test,  $p < 0.01$ , Figure S5), following the same profile as previously described for PSLMW and PSF solutions (Figure 1 and Figure S2, respectively). The  $R_{D/T}$  from PSHMW were also negatively correlated with the Corg in soils (Spearman,  $\rho = -0.85$ ,  $p < 0.01$ ). Compared to the  $R_{D/T}$  from PSLMW, those from PSHMW were overall higher, notably due to the higher amount of dimer being formed in the pyrolysis of PSHMW. The  $R_{D/T}$  from

PSHMW were not correlated with the sand and clay content of the matrices (Spearman,  $p > 0.05$ ).

Calibration curves were built from the pyrolysis of PSHMW powder spiked in different matrices ( $R^2$  greater than 0.75, Table S4). Both markers, dimer and trimer, were detected in the lowest concentration, i.e.,  $0.05 \text{ g kg}^{-1}$ , only in SA. For all the soil matrices, these markers were only detected in concentrations above  $0.10 \text{ g kg}^{-1}$ , equivalent to  $0.5 \mu\text{g}$  of polymer. The slopes of curves built with the styrene-dimer varied significantly with matrix type by about 3.0-fold. Specifically, the slope of the curve in GL was significantly smaller than DC, EP and SA (emmeans,  $p < 0.01$ , Figure S6a). For the trimer, the slopes of curves showed a larger variation up to 7.5-fold (Figure S6b), and GL soil presented a significantly steeper curve than DA, EP, and EC (emmeans,  $p < 0.01$ ). The slopes of curves built with the trimer were positively and strongly correlated with the Corg of matrices (Spearman test,  $\rho = 0.88$ ,  $p < 0.01$ ), similarly as observed for the curves built with this marker from PSLMW. No significant correlation was found between the slopes and the contents of sand and clay in the matrices (Spearman test,  $p > 0.05$ ).

## Discussion

We demonstrate in this work that the pyr-GC-MS analysis of PS is affected by external and internal factors interfering with the formation of its main markers. The first is a matrix effect, which affected the formation rate of markers and the slope of the calibration curves, and was circumvented by the normalization with PSF. The second, the polymer Mw, which directly affected the pyrolysates yields of PS and the slope of the calibration curves, leading to estimate errors in microplastic concentration.

Pyr-GC-MS analysis of PS microplastics, one of the most commonly used materials and largely found in the soil environment as debris (PlasticsEurope 2020; Huang et al. 2021), was strongly affected by the presence of matrices. Previous studies have shown an increased background in pyrograms with organic-rich soils (up to 5.6% Corg) mainly due to the detection of a PS marker, styrene, originating, for instance, from chitin or phenylalanine content (Fischer and Scholz-Böttcher 2017; Steinmetz et al. 2020). Here we showed that the Corg of soils was indeed strongly correlated with their styrene content naturally present, and none of the two other PS markers was detected. With the addition of PS in different matrices, the formation of its markers significantly differed, notably with a higher formation of trimer with the increasing Corg. A similar behavior was noticed during PSF pyrolysis, with higher formation of trimer- $F_3$  with the increasing Corg in matrices, highlighting the existence of matrix effect likely related to the Corg. The higher yield of trimer might be a consequence of reduced secondary reactions, preventing the conversion of trimer to dimer and/or monomer (Zhou et al. 2016), and here correlated to the greater presence of other organic compounds. Though the constituents of these soils were not fully characterized, no other significant or strong correlation was found between the other matrix parameters and the formation of markers. The presence of inorganic constituents, or their interaction with the organic ones, might also interfere in the PS pyrolysis (Fabbri, Trombini, and Vassura 1998; Bu et al. 2017). For instance, the adsorptive effect of clay materials was showed to cause a reduced detection of all PS markers (Fabbri, Trombini, and Vassura 1998; Bouzid et al.

2022). Clays might exhibit either a catalysis or a pyrolysis-inhibiting effect during pyrolysis, depending on the nature of the organic compound and of the clay-organic matter complex (Bu et al. 2017; Zhou, Yang, and Wang 2017). No correlation was found between the amount of clay in the soils and the formation of pyrolysis markers; however, the different natures of clay might play a role in the pyrolysis yields of microplastics (Faure et al. 2006). Further studies are necessary to fully elucidate the matrix components that interfere in the formation of polystyrene markers during the pyr-GCMS analysis and how they might affect the pyrolysis of other microplastic types.

We add evidence that the use of external calibration curves built in distinct matrices for the quantification of microplastic lead to great errors (Bouزيد et al. 2022). To overcome these matrix effects, the use of PSF as internal standard was tested; once this polymer exhibited similar properties to those of PSLMW and PSHMW against different matrices. The effect of different matrices on  $R_{D/T}$  was similar throughout the experiments performed with different polymers and methods of preparation, i.e., liquid solution of PSLMW and PSF or solid dilution of PSHMW. Therefore, the normalization of the analytes surface areas by those from PSF markers reduced the differences in the slopes of the calibration curves built in different matrices. This was only achieved when the corresponding fluoride marker was used, what is aligned with the principles of ISTD use, including sharing similar chromatographic properties (Unice, Kreider, and Panko 2012; Zhou, Yang, and Wang 2017).

An overall reduced formation of the three monitored pyrolysates was observed for PSLMW compared to PSHMW, leading to a considerable mass deficit. This might be attributed to the generation of other compounds unscreened for. For example, unsaturated compounds and heavier pyrolysates, i.e., oligomers with more than three repeating units, which are hardly analyzed by the common chromatographic-MS technique, might be preferably formed with the decreasing Mw (Costa and Camino 1982; Bouster, Vermande, and Veron 1989). Reduced yields of styrene dimer and trimer, to a negligible level, were also already observed for nanoplastics compared to microplastic, indicating a different formation or pyrolyzates from the microscale to the nanoscale (Blanco et al. 2021). The relative yields of PS markers significantly differed in the polymers with different Mw, notably the  $R_{D/T}$ , attributed to the higher quantity of dimer formed during the PSHMW pyrolysis. The thermo-degradation (or pyrolysis) of polymers are believed to follow a radical-based reaction, initiating mainly by random or end chain scission and forming macroradicals as stable as possible (Zhou et al. 2016; Duan et al. 2021). The initiation through chain-end scission might be particularly important with the decreasing of the Mw and the consequent increase in chain-end positions (Faravelli et al. 2001). The increase in chain-end positions in PSLMW, and likely the prevalence of an initiation reaction though chain-end scission, might play a role in the propagation reaction, thus privileging the formation of other markers instead of the dimer.

The formation of pyrolysates depends upon the microstructure of the polymer and the particle and on the relative bond strengths (Bouster, Vermande, and Veron 1989; Sam 2019; Park and Lee 2021; Miller et al. 2022). Changes in the aliphatic backbone of polymers as consequence of the weathering of microplastic in the environment, through mechanisms of chain scission and oxidation (Gewert, Plassmann, and MacLeod 2015;



651 Cai et al. 2018; Liu et al. 2021), might result in a lower  $M_w$  than the pristine microplas-  
652 tics used to build calibration curves. In this study, we present that such differences  
653 between the  $M_w$  led to quantification errors up to 5-fold, depending on the marker  
654 chosen. The consequences of the differences in polymer microstructure, such as degree  
655 of branching or cross-linking, on the quantification of polystyrene and other microplas-  
656 tics using pyr-GC-MS, remain to be investigated.

657 The PSHMW was insoluble in the solvents tested, likely due to its high  $M_w$  as the  
658 solubility and rate of dissolution of polymers are known to decrease with increasing  
659 molecular chains (Chojnacka, Janssen, and Schoenmakers 2014). Therefore, microplastic  
660 particles of PSHMW were added to solid matrices, which were mixed and milled. For  
661 the comparison with PSLMW, the same procedure was repeated for this polymer; i.e.,  
662 microplastic particles of a similar size distribution were added to sand and submitted to  
663 several milling processes. The heterogeneity of solid samples prepared through a milling  
664 and mixing method directly affect the deviation among triplicates and the linearity of  
665 the curves, as observed in this study, which are considerably lower than those obtained  
666 in solution. It is worth noting that the mechanical and visco-elastic properties of poly-  
667 mers usually rise with increasing  $M_w$  (Balani et al. 2015), likely representing a higher  
668 inhomogeneity of PSHMW compared to PSLMW samples. However, the relative stand-  
669 ard deviations between replicates and slopes of the curves were similar between these  
670 two polymer types. For the purposes of this study, the differences observed between the  
671 slopes of curves and the differences in the pyrolysates yields obtained from the two pol-  
672 ymers highlight the relevance of  $M_w$  differences in the quantification of microplastics.  
673 For studies aiming the quantification of microplastics in environmental samples, it is  
674 important to reduce the uncertainties related with calibration curves by performing the  
675 standard-addition method (Rodriguez et al. 1995; Cimetiere et al. 2013) or improving  
676 the sample preparation procedure.

677 The direct analysis of spiked soils using pyr-GC-MS, without further pretreatment  
678 besides drying, sieving, and grinding, allowed the detection of PS in a concentration  
679 above  $0.25 \text{ g kg}^{-1}$ , which was already reported in environmental samples, such as bio-  
680 solids (Okoffo et al. 2020; Ainali et al. 2021b; Zhang et al. 2022). Nevertheless, this limit  
681 of detection remains considerably high and might be even higher for polymers which  
682 generates hundreds markers of similar intensity, such as polypropylene and polyethylene  
683 (Tsuge, Ohtani, and Watanabe 2011; La Nasa et al. 2020). A direct analysis of complex  
684 matrices, rich in organic and inorganic components, might affect the method repeatabil-  
685 ity due to the overload of undesirable constituents. Due to the limited amount of sam-  
686 ple possible to be injected into the pyr-GC-MS, sample homogeneity is an important  
687 aspect (Dierkes et al. 2019; Steinmetz et al. 2020). Improving sample clean-up or pre-  
688 concentration through enzymatic and chemical digestion, density separation, or apply-  
689 ing pressurized liquid extraction, has been already proposed to increase the sensibility  
690 and/or remove matrices interferences (Fischer and Scholz-Böttcher 2017; Okoffo et al.  
691 2020; Thomas et al. 2020). In some instances, the treatment of the sample was insuffi-  
692 cient to eliminate this matrix effect (Bouzid et al. 2022), highlighting the importance of  
693 elucidating matrix effect previously to microplastic quantification. We argue that the  
694 well-known advantages of pyrolysis techniques, such as the minimal sample preparation,  
695 may be further explored to increase the sample purification. For example, the  
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701 application of multiple heating steps on each sample could allow the discrimination of  
702 the thermolabile natural organic matter previously to the pyrolysis of microplastics  
703 (Quénéa et al. 2006; Terán, Gonzalez-Vila, and Gonzalez-Perez 2009; Okoffo et al. 2020;  
704 La Nasa et al. 2021) and potentially contributing to higher sensitivity of microplastic  
705 detection in complex soil matrices.  
706

## 707 **Conclusion**

708 A robust quantification of PS using pyr-GC-MS through external calibration curves  
709 must include the elucidation of potential matrix effects, circumventing them by the use  
710 of an internal standard, and the use of a reference standard that shares a similar Mw  
711 range. Without prior knowledge of the Mw, efforts must be made to estimate quantifi-  
712 cation errors and incorporate measurement uncertainties. In most cases, an underesti-  
713 mation of microplastic concentration is expected, given the potential weathering of the  
714 plastic found in the environment, which encompass molecular chain scission and, con-  
715 sequently, reduced Mw. The formation of styrene-trimer was less affected when pyrolyz-  
716 ing PS of different Mw and this marker is, therefore, recommended for the  
717 quantification of PS microplastic in samples. Finally, the direct analysis of soils enriched  
718 with PS, without any pretreatment, resulted in the detection of PS markers at concen-  
719 trations already observed in the environment. However, further sample preparation  
720 efforts are needed to improve the robustness and sensitivity of microplastic quantifica-  
721 tion using pyr-GC-MS  
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726

## 727 **Disclosure statement**

728 There are no relevant financial or non-financial competing interests to report.  
729

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