

Introduction

Glioblastoma (GBM)
Most aggressive brain tumor
12-18 months median survival

Blood-brain-barrier (BBB)

- Protects the brain from most external aggressions
- Allows the passage of nutrients, oxygen and metabolic waste

KNOWLEDGE GAP

SIBorG project
A vascularized GBM-on-chip model

New engineered tool using organ-on-chip technology

Functional vascularization:

- ❖ Complex and perfused microvascular architecture
- ❖ Dynamic conditions to mimic the blood flow
- ❖ GBM cells to recreate GBM microenvironment

Relevant alternative

- ❖ Bioengineering tool for drug nano-vectorization
- ❖ Alternative to animal testing

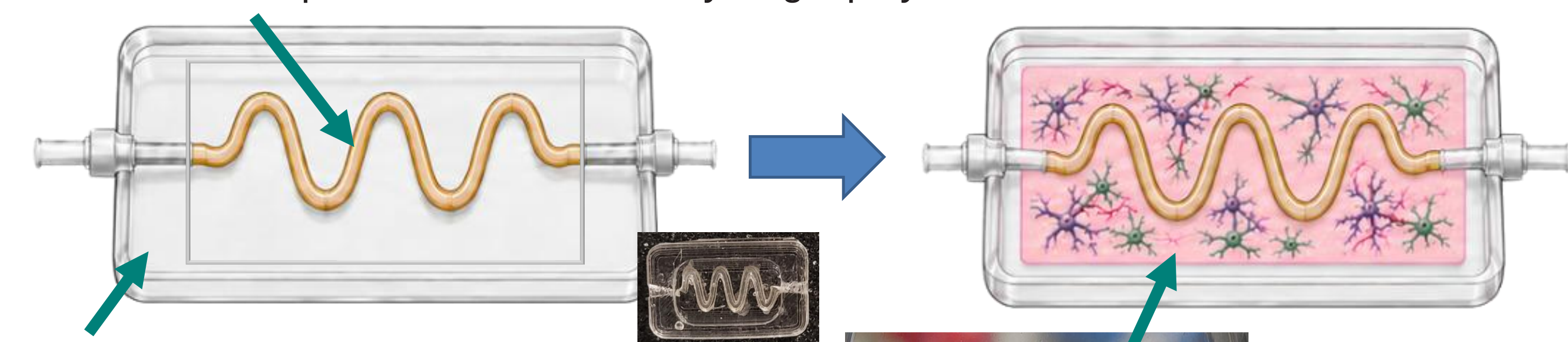
Integrated monitoring

- ❖ Real-time detection of nanocarriers transport, pH, O₂
- ❖ BBB integrity, therapeutic delivery and hypoxia

BBB-on-chip device

First complex design as vascular architecture

- Ongoing COMSOL Multiphysics® simulation to determine the optimal vascular design
- Sacrificial template, dissolved after hydrogel polymerization for channel formation



PDMS mold:

- Cast inside central compartment around the sacrificial template

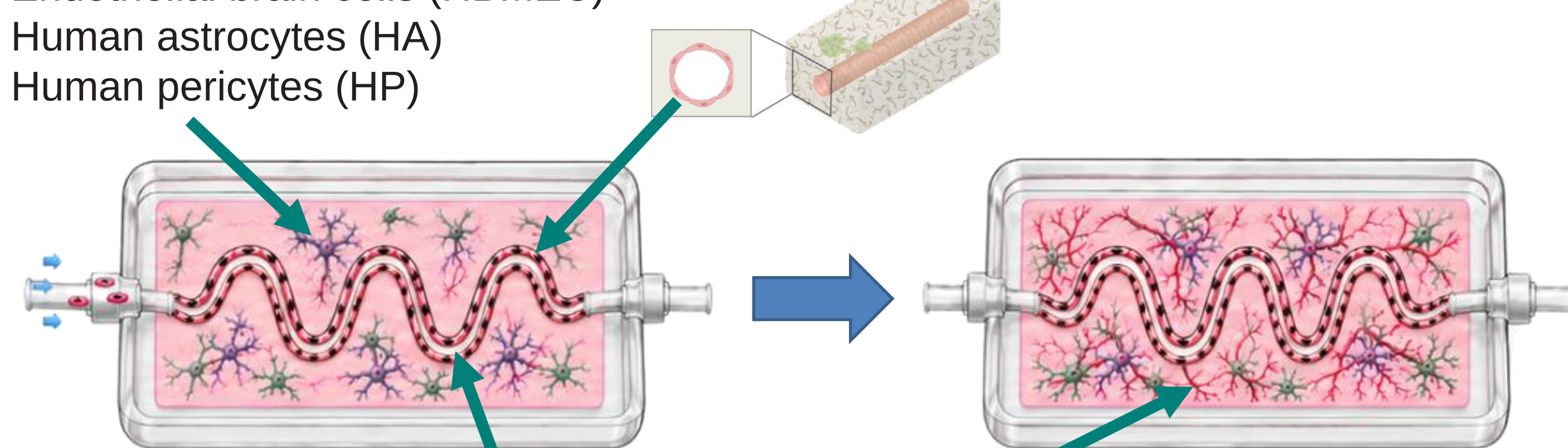
Hydrogel as 3D scaffold

- 3D cell organization and interactions
- Extracellular matrix protein
- Pooled inside central compartment around sacrificial template

3D cell culture:

- Endothelial brain cells (HBMEC)
- Human astrocytes (HA)
- Human pericytes (HP)

Template-Based



MACRO and MICRO-vascularization:

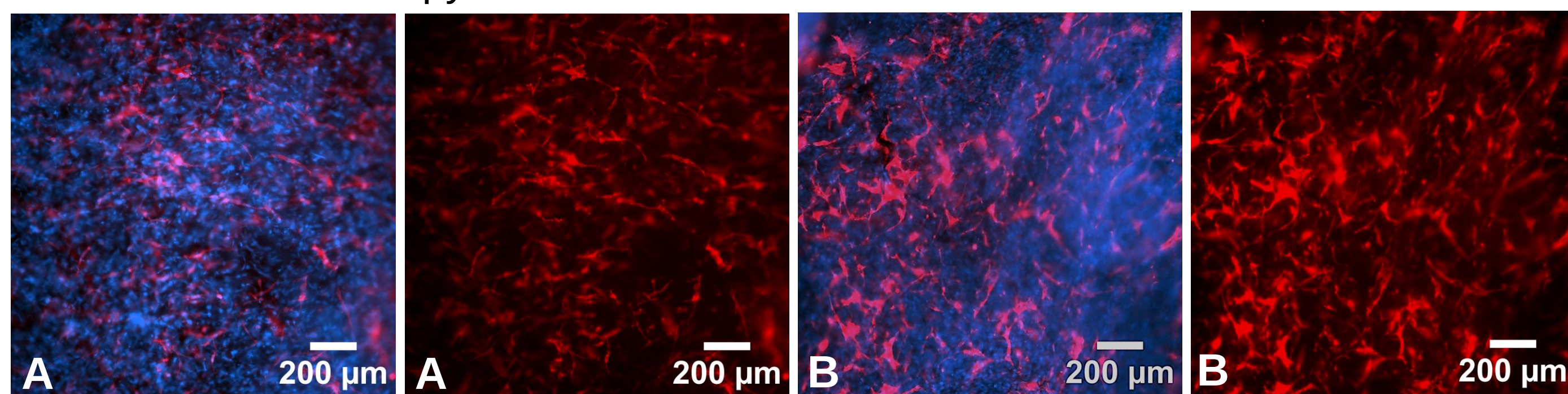
- Gelatin-based arteriole template used as a sacrificial material
- Self-assembled capillaries through angiogenesis

Device optimization

1 Hydrogel composition : thrombin, fibrinogen and collagen-I

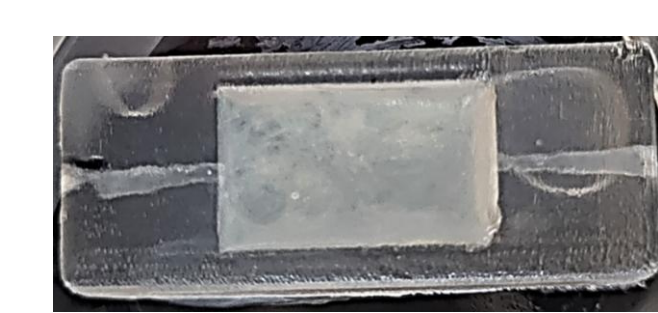
Tests:

- 8 different concentrations, hyaluronic acid addition
- Physicochemical and rheological properties
- Confocal microscopy observation : endothelial network formation



Endothelial cells after 7 days (Hoechst) and endothelial network formation (CD31) with 2.5 mg/mL (A) or 5 mg/mL (B) hyaluronic acid

→ final optimal hydrogel: thrombin (1 U/mL), collagen-I (2.5 mg/mL), fibrinogen (5 mg/mL) and hyaluronic acid (2.5 mg/mL)



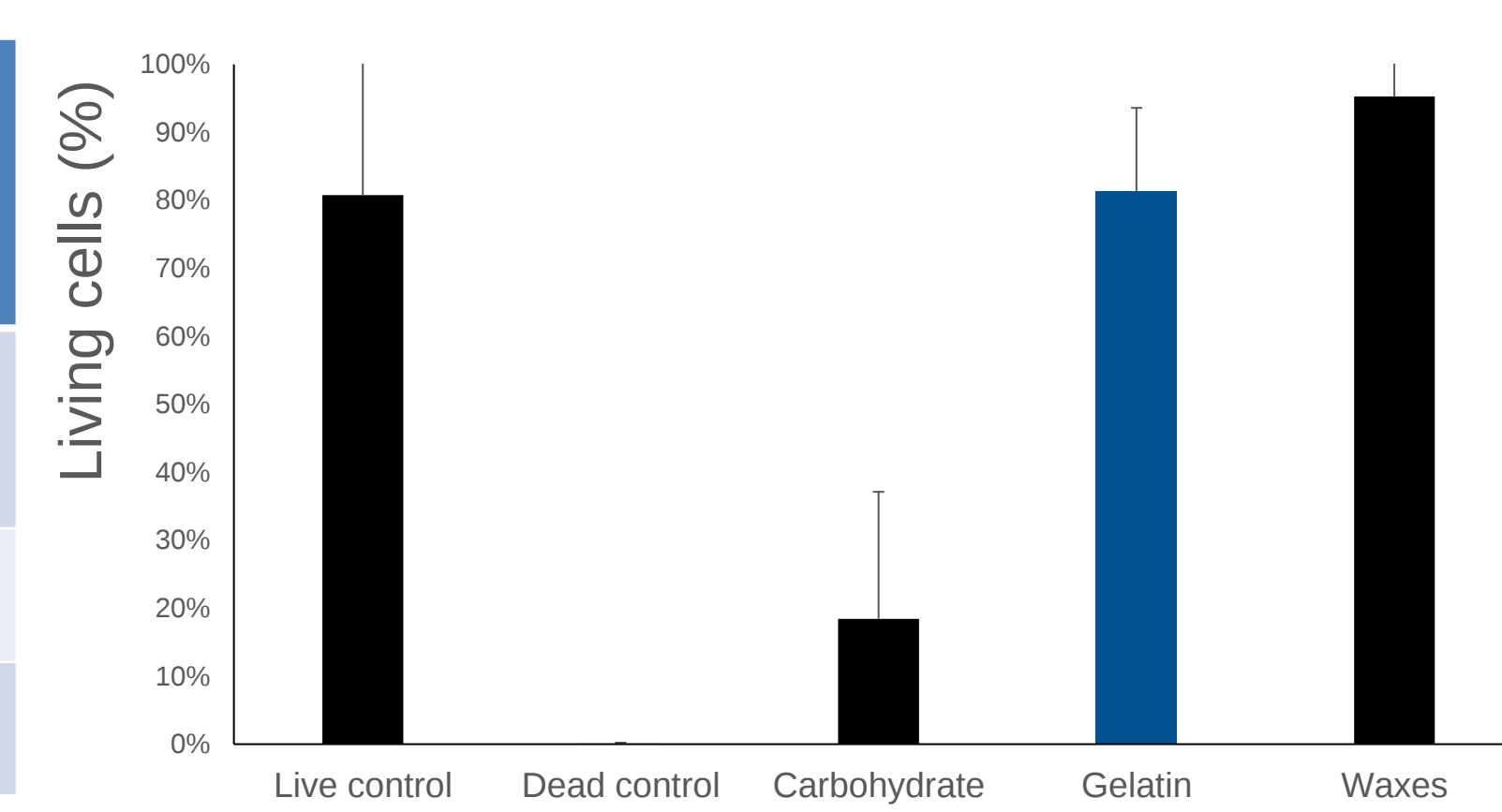
Hydrogel after 7 days
CD31
Hoechst

2 Sacrificial material:

3 materials evaluated: carbohydrate glass, gelatin and paraffin/polyester wax

- ✓ Physicochemical properties
- ✓ Biocompatibility on cells (live dead protocol)

	Synthesis	Molding demolding in PDMS	Dissolution at 37°C
Carbohydrate glass	✓	✗	✓
Gelatin	✓	✓	✓
Waxes	✓	✓	✗



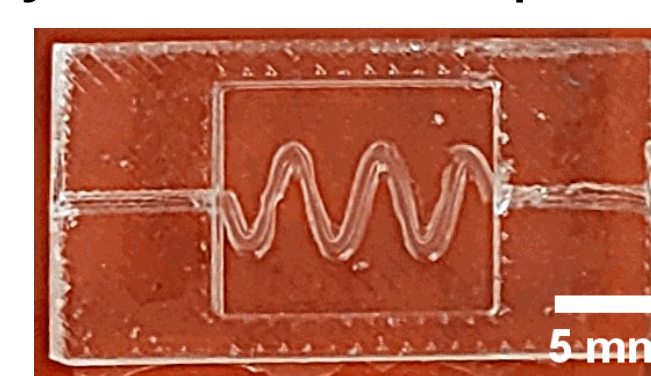
Physicochemical properties

Biocompatibility on HBMEC

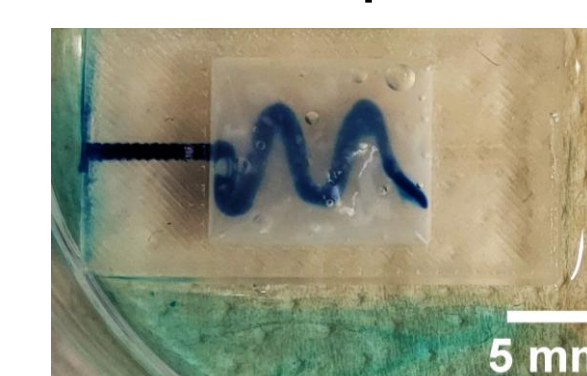
→ validation of gelatin as optimal sacrificial material

Channel formation:

- Insertion of a gelatin template into the PDMS microchip
- ongoing assays with 3D bio-printing to obtain well-defined template



Microchip support with gelatin template



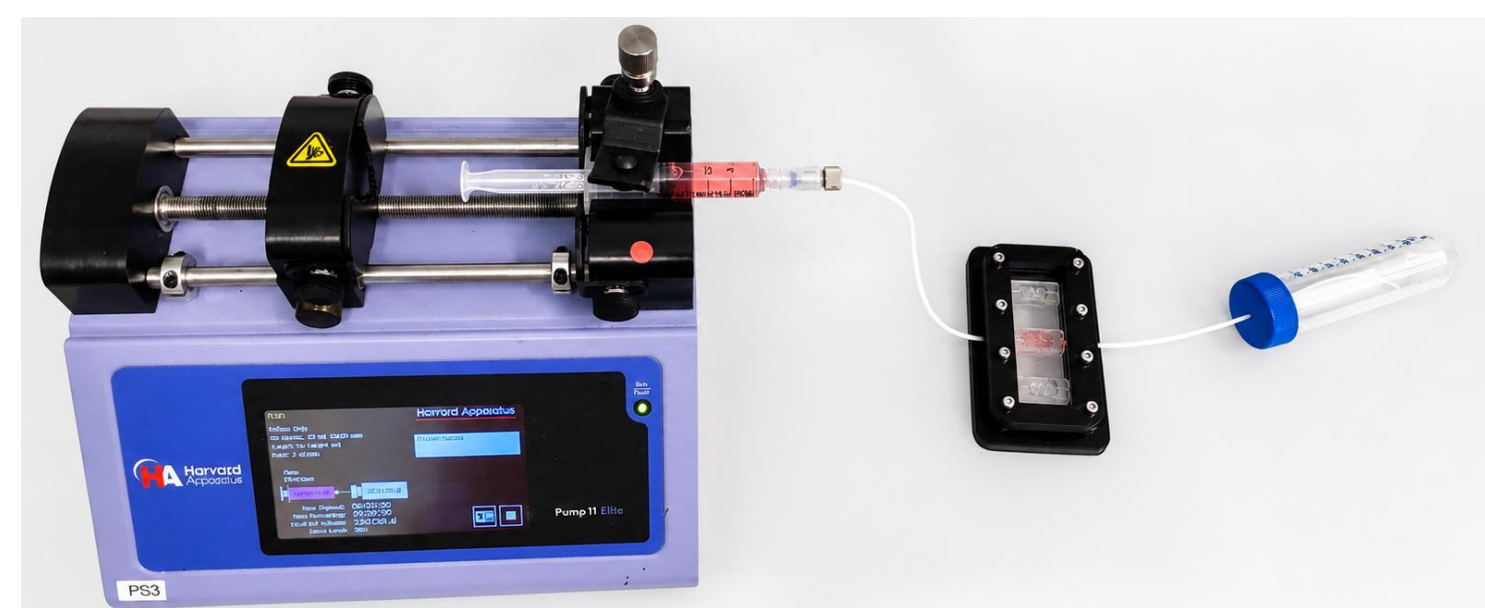
Microchip support after gelatin dissolution

3 Culture under perfusion:

- ✓ Initial perfusion experiments are ongoing
- ✓ The chip must be closed during culture for a stable flow
- ✓ Reversible support system "Lock and Play" (using CAD and 3D printing) to ensure closure



Lock and Play device



Dynamic culture setup

Perspectives:

- ✓ Ongoing work to determine the optimal vascular architecture regarding shear stress
- ✓ Integration of sensors for real-time monitoring of O₂ and pH
- ✓ Validation of the model with integrity and permeability assays, detection of nanocarriers
- ✓ Incorporation of GBM cells to establish a GBM-on-chip model

Conclusion

- Gelatin is an ideal sacrificial material for complexifying vascular architecture of the BBB-on-chip device due to its favorable physicochemical and biocompatible properties
- Ongoing COMSOL Multiphysics® simulations aim to determine the optimal design taking into account shear stress constraints
- O₂ and pH sensors will allow real-time metabolic monitoring, while BBB integrity and permeability will be evaluated
- Incorporating U87 cells will establish a GBM-on-chip model

Our model provides a promising platform to investigate nanocarrier transport and evaluate drug delivery strategies in a physiologically relevant microenvironment

References: Figarol *et al.*, 2021, Biomedical Materials 16, n° 1: 015006, <https://doi.org/10.1088/1748-605X/aba5f1>
Golden *et al.*, 2007, Lab on a Chip 7, n° 6: 720, <https://doi.org/10.1039/b618409j>
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